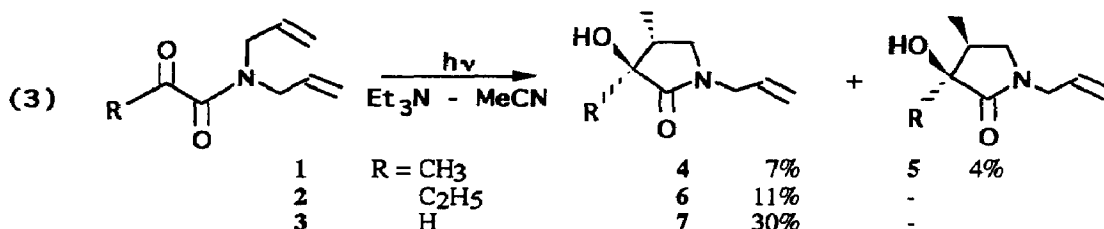
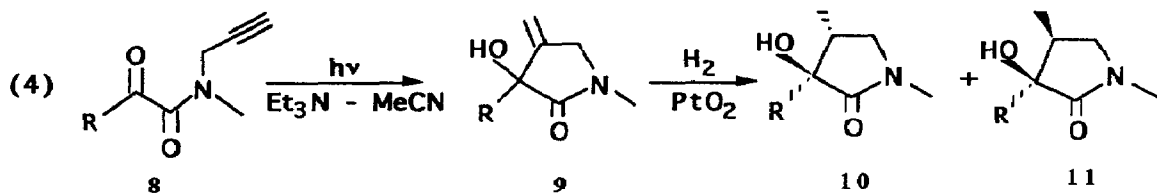


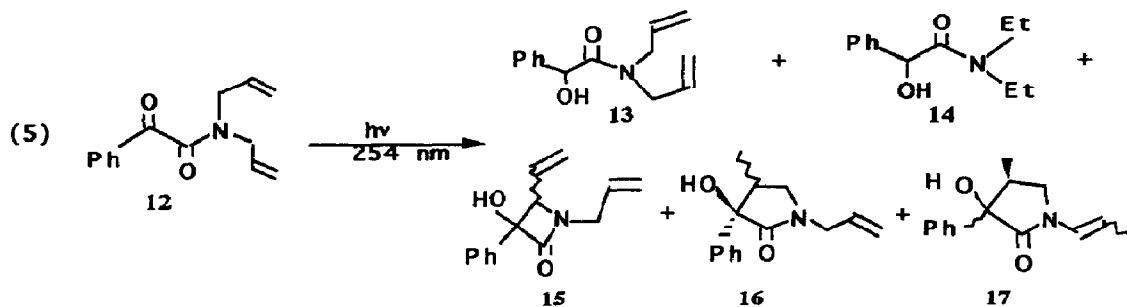
When solutions of  $\alpha$ -ketoamides 1 or 2 and triethylamine were irradiated in acetonitrile ( $10^{-2}$ M) at 254 nm, the hydroxypyrrolidones 4-6 were isolated in low yields from the complex reaction mixture (equation 3). Similar results are obtained when the irradiation was carried out in HMPA and  $\text{Et}_3\text{N}$  (10 equivalents).



When the irradiation of aldehyde 3 was carried out under similar conditions, 7 was isolated as the only butyrolactam. Similarly, the propargyl ketoamide 8 led to the lactam 9 (50%). The stereochemistry of the lactams 4 and 5 was established by comparison of their NMR spectra<sup>7</sup> with those of the lactams 10 and 11 obtained stereoselectively by hydrogenation of the allylic alcohol 9.<sup>8</sup>



The behaviour of the benzoylformamide 12 under photoreductive conditions depends on the nature of the reducing agent (equation 5, Table). In pure HMPA the benzylic alcohol 13 and the  $\beta$ -lactam 15 arising from a  $\gamma$ -hydrogen abstraction process, could be isolated. No  $\gamma$ -lactam could be detected under these conditions.

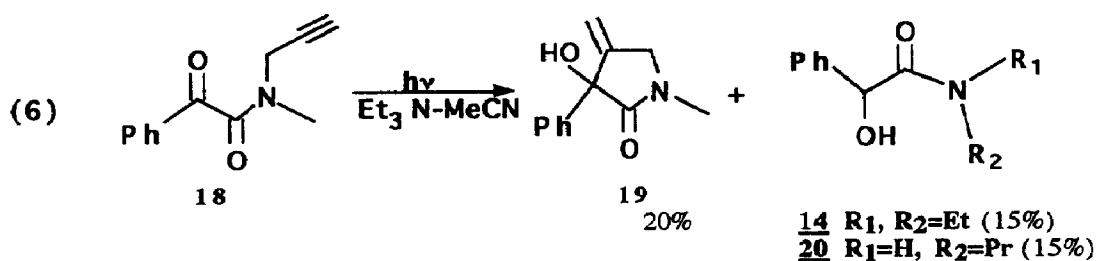


**Table :** Yields (%) of the products isolated from the irradiation of **12** at 254 nm.

Solvent	<b>13</b>	<b>14</b>	<b>15</b>	<b>16</b>	<b>17</b>
HMPA	18%	-	30%	-	-
Et <sub>3</sub> N-MeCN	-	20	22	20	10
Et <sub>3</sub> N-HMPA	-	20	7	25	-

When the irradiation of **12** was conducted in the presence of triethylamine, the benzylic alcohol **14** was preferred to **13**, and the  $\gamma$ -lactam **16** was preferred to the  $\beta$ -lactam **15** when the photolysis was carried out in a solution of triethylamine in HMPA ( $10^{-2}$ M).

The photolysis of the propargylamide **18** in a solution of Et<sub>3</sub>N (10 equivalents) in MeCN led to a mixture of the  $\gamma$ -lactam **19** and the benzylic alcohol **14**.



Production of  $\gamma$ -lactams by photolysis of  $\alpha$ -ketoamides in a reducing medium contrasts with the results described for  $\alpha$ -ketoesters and is in agreement with recent results on the ring closure of  $\alpha$ -alkoxyester free radicals <sup>5c</sup>.  $\gamma$ -Lactams could result from an intermolecular electron transfer from the reducing agent to the excited carbonyl group and consequently from an anion radical intermediate. The replacement of an alkyl group by a phenyl substituent on the  $\alpha$ -oxoamide stabilizes the  $\alpha$ -hydroxyamide free radical intermediate by delocalisation and allows competitive processes to occur. Formation of products **15** derived from a Norrish type II process and benzylic alcohols **13** or **14** now compete with  $\gamma$ -lactam production. The formation of benzylic alcohol **14** might be due to diethylamine present in very small amounts in the triethylamine used and its addition to the hydroxyketene expected from the Norrish type II elimination of **12**. To test this hypothesis we carried out the irradiation of **12** in a mixture of triethylamine and acetonitrile in the presence of *n*-propylamine. Besides **15** and **17** we now isolated the *N*-methyl, *N*-propyl mandelamide **20** (15%) rather than the corresponding diethylamide **14**.

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- 7 Selected NMR data  $\delta$  (ppm) in  $\text{CDCl}_3$ . **4**:  $^1\text{H}$  NMR:  $\delta$  1.09 (d, 3H,  $J=7$  Hz), 1.20 (s, 3H), 1.85 (sl, 1H), 2.38 (m, 1H), 2.78 (dd, 1H,  $J=9.5$  and 9.4 Hz), 3.33 (dd, 1H,  $J=9.5$  and 8 Hz), 3.87-4.05 (m, 2H), 5.14-5.24 (m, 2H), 5.64-5.78 (m, 1H);  $^{13}\text{C}$  NMR:  $\delta$  11.60 (q), 18.83 (q), 39.14 (t), 45.37 (t), 49.50 (d), 76.09 (s), 118.28 (t), 131.81 (d), 177.26 (s). **5**:  $^1\text{H}$  NMR:  $\delta$  1.08 (d, 3H,  $J=7$  Hz), 1.37 (s, 3H), 1.83 (sl, 1H), 2.15 (m, 1H), 2.97 (dd, 1H,  $J=9.7$  and 5.7 Hz), 3.33 (dd, 1H,  $J=9.7$  and 7 Hz), 3.8-4.05 (m, 2H), 5.14-5.24 (m, 2H), 5.64-5.78 (m, 1H);  $^{13}\text{C}$  NMR:  $\delta$  12.11 (q), 23.48 (q), 38.18 (t), 45.40 (t), 50.90 (t), 77.19 (s), 118.18 (t), 131.84 (d), 177.0 (s). **9**:  $^1\text{H}$  NMR:  $\delta$  1.25 (s, 1H), 1.45 (s, 3H), 2.95 (s, 3H), 3.93 (dt, 1H,  $J_1=14$ ,  $J_2=1.9$  Hz), 4.0 (dt, 1H,  $J_1=14$ ,  $J_2=1.9$  Hz), 5.20 (t, 1H,  $J=1.9$  Hz), 5.47 (t, 1H,  $J=1.9$  Hz);  $^{13}\text{C}$  NMR:  $\delta$  25.60 (q), 29.62 (q), 51.47 (t), 73.71 (s), 109.69 (t), 144.94 (s), 175.52 (s). **16a**:  $^1\text{H}$  NMR:  $\delta$  1.08 (d, 3H,  $J=7.1$  Hz), 1.72 (sl, 1H), 2.37-2.48 (m, 1H), 3.07 (dd, 1H,  $J=9.8$  and 6.7 Hz), 3.37 (dd, 1H,  $J=9.8$  and 7 Hz), 3.93-4.03 (m, 2H), 5.18-5.29 (m, 2H), 5.68-5.86 (m, 1H), 7.20-7.40 (m, 5H);  $^{13}\text{C}$  NMR:  $\delta$  11.35 (q), 40.83 (t), 45.72 (t), 50.89 (d), 80.28 (s), 118.45 (t), 125.58 (d), 126.96 (d), 127.70 (d), 128.31 (d), 131.94 (d), 142.27 (s), 174.79 (s). **16b**:  $^1\text{H}$  NMR:  $\delta$  0.67 (d, 3H,  $J=6.7$  Hz), 1.28 (sl, 1H), 2.6-2.7 (m, 1H), 2.86 (dd, 1H,  $J=9.8$  and 9.4 Hz), 3.39 (dd, 1H,  $J=9.4$  and 8.6 Hz), 4.05 (d, 2H,  $J=6.3$  Hz), 5.25-5.38 (m, 2H), 5.8-5.95 (m, 1H), 7.2-7.4 (m, 5H);  $^{13}\text{C}$  NMR:  $\delta$  12.96 (q), 40.48 (d), 45.66 (t), 49.77 (t), 81.22 (s), 118.60 (t), 125.56 (d), 127.72 (d), 128.23 (d), 131.69 (d), 138.83 (s), 175.62 (s).
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